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ARE THE OPTIMUM PHARMACEUTICAL PREPARATIONS USED FOR THE SECOND-LINE ERADICATION THERAPY FOR HELICOBACTER PYLOR/INFECTION IN JAPAN? - A DISCUSSION FROM A SIMULATION FOR THE AMOUNT OF ANTIBIOTICS IN STOMACH BASED ON THE DATA OF DISSOLUTION STUDIES -

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ABSTRACT: The purpose of this study was to evaluate whether the optimum preparations are used for the second-line eradication therapy for Helicobacter pylori (H. pylori) infection in Japan. In the therapy, commercial tablets, which have 250 mg of amoxicillin (AMX) and 250 mg of metronidazole (MTZ), are used as antibiotics. The evaluation was performed by dissolution tests and simulations for intragastric drug concentrations based on the dissolution data. The dissolution tests were performed using the paddle method of Japanese pharmacopoeia XV. The data obtained were used for the simulation of intragastric drug concentrations. In dissolution studies, the 100% dissolution time (T100%) of AMX from the tablet which was about 20 min was faster than that from AMX alone (about 6 h). $\rm T_{100\%}$ of MTZ from the tablet which was about 3 h was later than that from MTZ alone (about 30 min). In simulated intragastric concentrations, the time above minimum inhibitory concentration (MIC) of the AMX tablet was 6.6 h which was shorter than that of AMX (11.4 h). The Cmax of the MTZ tablet was 0.095 mg/ml which was lower than that of MTZ (0.190 mg/ml). However, AMX is a timedependent antibiotic, longer duration above MIC is desirable. On the other hand, MTZ is a concentration-dependent antibiotic, higher Cmax is desirable. In conclusion, the commercial products are found to be not the best, and preparing an AMX pharmaceutical preparation with more sustained release characteristic and a MTZ preparation with more rapid release characteristic are considered to be very advantageous.

KEYWORDS: : Amoxicillin; Metronidazole; *Helicobacter pylori*; Dissolution test; Simulation

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ABBREVIATIONS

AMX – Amoxicillin CAM – Clarithromycin FLG – Flagyl H. pylori – Helicobacter pylori JP – Japanese pharmacopoeia MTZ – Metronidazole MIC – Minimum inhibitory concentration PST – Pasetocin Tablets 250 PPI – Proton pump inhibitor TAM – Time above MIC

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INTRODUCTION

astric Helicobacter pylori (H. pylori) infection is associated with chronic (type B) gastritis and peptic ulcer [1-3]. H. pylori are often observed to adhere to the antral epithelium of the human stomach and gastric metaplasia in the duodenum. Gastric and duodenal ulcers are believed to develop as the result of damage to the gastric mucosa by cytotoxic substances (ammonia, cytotoxin, etc.) produced by H. pylori [4]. In developed countries, at least 20% of people younger than 20 years old and 50% of those older than 60 years old are infected [5]. In developing countries, the rate of infection is much higher, with a prevalence of 85% in northern Nigeria and 71-79% in other African countries [6]. Even if gastric and duodenal ulcers with *H. pylori* infection are treated, it will most likely cause the recurrence of the ulcers [7-9]. Therefore, the eradication of *H. pylori* is considered to be very important.

The primary eradication therapy for *H. pylori* in Japan which is a triple therapy using amoxicillin (AMX), clarithromycin (CAM) and proton pump inhibitor (PPI) was started in 2000 with the therapy approved for health insurance coverage (10). However, because of the increase of incomplete eradication, the second-line eradication therapy was approved in 2007 (10). In the eradication therapy, metronidazole (MTZ) is used in the place of CAM. The *H. pylori* strains with CAM resistance significantly increase more and more, so the importance of the second-line eradication therapy also increases [10].

On the other hand, the pharmaceutical preparations used in the eradication therapy in Japan were not developed for the eradication therapy. For example, Pasetocin Tablets 250 (PST) for AMX and Flagyl (FLG) for MTZ were developed in 1981 and 1963, respectively, for various bacterial infections. From the point of view of the design of pharmaceutical preparations, there is a question whether the used preparations are the optimum or not, because these preparations were designed to exert the effect through the blood circulation after absorbed from the GI tract. For *H. pylori* eradication therapy, a direct attack in stomach is expected [11].

Therefore, we selected the commercial pharmaceutical preparations used in second-line therapy due to the increasing importance, and performed the simulation studies for the intragastric concentrations of the antibiotics, AMX and MTZ from the results of dissolution test. And we tried to evaluate whether they are optimal preparations or not.

MATERIALS AND METHODS Materials

Amoxicillin (AMX) and metronidazole (MTZ) were purchased from Sigma Chemical Company (St. Louis, MO, USA) and Wako Pure Chemical Industries, Ltd. (Osaka, Japan), respectively. Pasetocin Tablets 250 (PST) and Flagyl (FLG) which were from Sigma Pharmaceuticals Pty. (Croydon, Australia) and Shionogi Pharmaceutical Co. (Osaka, Japan) were used as the pharmaceutical preparations for AMX and MTZ. Other chemicals were of special reagent grade or HPLC grade.

Dissolution test

Dissolution characteristics of AMX and MTZ, and those tablets were evaluated by the paddle method of Japanese pharmacopoeia (JP) XV at 50 rpm of paddle speed using JP XV apparatus. The media used were 900 ml of the first and the second fluids of JP XV (pH 1.2 and 6.8, respectively), and pH 4.0 buffer solution, which were maintained at $37.0 \pm 0.5^{\circ}$ C. The buffer solution at pH 4.0 was made by mixing the first and the second fluids. The first fluid (pH 1.2) contains 2.0 g of NaCl and 7 ml of HCl in 1000 ml and the second fluid (pH 6.8) contains 250 ml of 0.2 M KH₂PO₄ and 118 ml of 0.2 M NaOH in 1000 ml. One milliliter of sample solution was taken at appropriate time intervals and assayed by HPLC. Fresh medium was added to replace the sample taken.

The Dissolution characteristics were determined for a 750 mg of AMX and 3 tablets of PST, and for a 250 mg of MTZ and a table of FLG.

Determination of AMX and MTZ by HPLC

The HPLC system consisted of a Shimadzu model LC-10AS pump, SIL-10A auto-injector, SPD-10A UV photometrical detector, and CTO-10A column oven equipped with a Shimadzu model SCL-10A system controller and C-R7A Chromatopac, all from Shimadzu Corporation (Kyoto, Japan). The chromatographic column was an YMC Pack AM312 ODS $(150 \times 6 \text{ mm, i. d.})$ obtained from YMC Co., Ltd. (Kyoto, Japan). The conditions for the flow rate, the wavelength for determination, and the temperature of the column were 1 mL/min, 254 nm, and 40°C, respectively. The mobile phase used for the determination of AMX and MTZ were acetonitrile-water, 1:19, V/V and 1:9, V/V. The retention times of AMX and MTZ were about 8.6 min and 4.9 min. The concentrations of AMX and MTZ were calculated using calibration curves for the antibiotics. The injection samples for HPLC were prepared by the following procedure: a 30 µl of sample solution was added to 270 µl of the second fluid of JP XV, and then mixed well. A 10 μ l of the solution was injected to HPLC.

The simulation of intragastric concentrations

Drug concentrations in stomach were simulated as shown in Fig. 1, using equations (1), and (2) based on the result of dissolution test. Figure 1: An image of simulation about equation (1) and (2). t_n is sampling time at n times (n is natural number), C_n is concentration at t_n in the dissolution test, X'_n is simulated intragastric quantity at t_n .



$$Ln X_{(n, min)} = -1.5379 (t_{n} t_{n-1}) + Ln X_{(n-1, max)}$$
(1)

$$X_{(n, max)} = X_{(n, min)} + (C_n - C_{n-1}) \times 900$$
 (2)

where X_n is simulated intragastric quantity at t_n and $X_{(0, max)} = X_{(0, min)} = 0$.

where t_n is sampling time at n times and n is natural number.

where C_n is concentration at t_n in the dissolution test and $C_0 = 0$.

In equation (1) and Fig. 1(a), it showed that as the result of intragastric emptying during $t_n \cdot t_{n-1}$, $X_{(n-1, max)}$ became $X_{(n, min)}$. The intragastric emptying rate of liquid is expressed as first-order kinetics, and a first order rate constant, 1.5379 is the emptying rate constant reported by Camilleri et al [12].

But equation (1) doesn't include further dissolution quantity between t_n and t_{n-1} . Therefore, we corrected it by using equation (2) and Fig. 1 (b). $(C_n \cdot C_{n-1}) \times 900$ is further dissolution between t_n and t_{n-1} in dissolution test. 900 is the volume of the test media (ml). It is thought that the dissolution quantity of AMX and MTZ in the dissolution test is same as in the stomach because of their solubility. Therefore, we thought that $(C_n \cdot C_{n-1}) \times 900$ is further dissolution quantity between t_n and t_{n-1} in stomach. At the end, we converted X into simulated intragastric concentration by multiplied X by 900 ml (stomach volume). In case of AMX, we calculated time above minimum inhibitory concentration (time above MIC, TAM) because AMX is time-dependent antibiotic, and in case of MTZ, we calculated Cmax because it is concentration-dependent antibiotic.

RESULATS AND DISCUSSION

Dissolution characteristics of AMX and MTZ

Fig. 2 (a) and (b) show the dissolution characteristics of AMX from 3 PST tablets and a 750 mg of AMX, respectively. In Fig. 2, the dissolution characteristics of pH 4.0 and 6.8 were almost the same. The dissolution rate of AMX at pH 1.2 showed more rapid than those at pH 4.0 and 6.8, which decreased over 30 min due to the degradation of AMX in acidic conditions [13]. On the eradication therapy, PPI was used together. Therefore, the dissolution pattern in stomach was considered to be equivalent to the pattern of the dissolution tests at pH 4.0 and 6.8. We employed the data at pH 4.0 and 6.8. The 100% dissolution time ($T_{100\%}$) of PST at pH 4.0 and 6.8, which were about 20 min, was faster than those of AMX which were about 6 h. Mai Kubodera et al.

Figure 2: In vitro dissolution profiles in ph 6.8 (•), 4.0 (\Box), and 1.2 (\blacktriangle). (a) AMX from 3 PST tablets, (b) AMX from a 750 mg of itself. Each point represents the mean \pm S.D. of 3 experimental runs.



The dissolution characteristics of AMX from a PST tablet and a 250 mg of AMX were confirmed to be almost same as the results in Fig. 2 (a) and (b) (data not shown).

Figures 3 (a) and (b) show dissolution characteristics of MTZ from a FLG tablet and 250 mg of MTZ, respectively. In Fig. 3 (a), the dissolution characteristic of pH 1.2 was rapid those of pH 4.0 and 6.8, and there was no difference between the results at pH 4.0 and 6.8. This difference between pH 1.2 and pH 4.0 and 6.8 might be caused by the sugar-coating layer of the preparation. The dissolution data at pH 4.0 and 6.8 were employed by the same reason in the case of AMX. The 100% dissolution time ($T_{100\%}$) of FLG which were about 3 h was later than those of MTZ which were about 30 min.

Figure 3: In vitro dissolution profiles in pH 6.8 (•), 4.0 (\square), and 1.2 (\blacktriangle). (a) MTZ from a FLG tablet, (b) MTZ from a 250 mg of itself. Each point represents the mean \pm S.D. of 3-6 experimental runs.



The simulation of intragastric concentrations Figures 4 (a) and (b) show intragastric simulated concentrations of AMX from PST and itself, respectively. The point which y is 0.0001 mg/ml is the minimum inhibitor concentration (MIC)

of AMX for the bacteria. Solid and dotted lines

are intragastric simulated concentration obtained

from dissolution test data at pH 6.8 and 4.0, re-

spectively. AMX is time-dependent antibiotic, so an efficacy depends on a length of time above MIC (TAM). Therefore, TAM was calculated from the mean value of dissolution test data at pH 6.8 and 4.0. The TAM values of PST and AMX were 6.6 h and 11.4 h, respectively. The value of PST was about three-fifths compared with that of AMX.

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Figure 4: Simulated intragastric behaviors in pH 6.8 (a solid line) and 4.0 (a dotted line). (a) AMX from 3 Pasetocin tablets, (b) AMX from a 750 mg of itself.

Figure 5: Simulated intragastric behaviors in pH 6.8 (a solid line) and 4.0 (a dotted line). (a) MTZ from a FLG tablet, (b) MTZ from a 250 mg of itself.



Figures 5 (a) and (b) show simulated intragastric concentrations of FLG and MTZ. MTZ is concentration-dependent antibiotic, so an efficacy depends on a height of maximum concentration (Cmax). Therefore, Cmax was calculated from the mean values of dissolution test data at pH 6.8 and 4.0. The values for FLG and MTZ were 0.095 mg/ml and 0.190 mg/ml, respectively. The Cmax of FLG was about a half compared with that of MTZ.

CONCLISION

It was found from our studies that the commercial products, both PST and FLG, were required to improve the drug release rate. In the case of PST, the simulated TAM value (about 6.6 h) was about three-fifths of AMX (11.4 h). The efficacy of

AMX depends on a length of TAM. It was found that the efficacy of PST might be poorer than that of AMX. In the case of FLG, the simulated Cmax value (about 0.095 mg/ml) was a half of MTZ (about 0.190 mg/ml). The efficacy of MTZ depends on a value of Cmax. It was found that the efficacy of FLG might be poorer than that of MTZ. In addition, our intragastric simulated concentrations were almost same as the clinical data [14].

Consequently, for eradicating *H. pylori*, preparing more sustained release preparations of AMX and more rapid release preparation of MTZ are considered to be very advantageous in order to decrease doses and to increase the efficacy over the commercial products. Journal of Basic and Clinical Pharmacy

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