

www.jbclinpharm.com



Antioxidant, Antihyperlipidaemic and Antidiabetic Activity of *Eugenia Floccosa* Bedd Leaves in Alloxan Induced Diabetic Rats

Mary Jelastin Kala S¹, Tresina P.S.² and Mohan V.R.^{2*}

¹ Department of Chemistry, St. Xavier's College, Palayamkottai, Tamil Nadu, India.

² Ethnopharmacology Unit, Research Department of Botany, V.O.Chidambaram College, Tuticorin-628008, Tamil Nadu, India.

ABSTRACT

The ethanol extract of *Eugenia floccosa* Bedd (Family: Myrtaceae) leaf was investigated for its antioxidant, antihyperlipidaemic and antidiabetic effect in Wistar Albino rats. Diabetes was induced in Albino rats by administration of alloxan monohydrate (150mg/kg, i.p.). The ethanol extracts of *E. floccosa* at a dose of 150 and 300mg/kg of body weight were administered at single dose per day to diabetes induced rats for a period of 14 days. The effect of ethanol extract of *E. floccosa* leaf extract on blood glucose, plasma insulin, creatinine, glycosylated haemoglobin, urea serum lipid profile [total cholesterol (TR), triglycerides (TG), low density lipoprotein – cholesterol (LDL-C), very low density lipoprotein – cholesterol (VLDL-C), high density lipoprotein – cholesterol (HDL-C) and phospholipid (PL)] serum protein, albumin, globulin, serum enzymes [serum glutamate pyruvate transaminases (SGOT), and alkaline phosphatase (ALP)], lipoprotein peroxidation (LPO) antioxidant enzymes (catalase (CAT), superoxide dismutase (SDD), reduced glutathione (CSH) and glutathione peroxidase (GPx) were measured in the diabetic rats. The ethanol extract of *Eugenia floccosa* leaf elicited significant reductions of blood glucose (P<0.05), lipid parameters except HDL-C, serum enzymes and significantly increased HDL-C and antioxidant enzymes. The extracts also caused significant increase in plasma insulin (P<0.05) in the diabetic rats. From the above results, it is concluded that ethanol extract of *Eugenia floccosa* possesses significant antidiabetic, antihyperlipidaemic and antioxidant effects in alloxan terms.

KEY WORDS

Antioxidant, antihyperlipidaemic, antidiabetic, E. floccosa, alloxan.

received on 16-12-2011 accepted on 02-08-2012 available online 15-02-2012 www.jbclinpharm.com

INTRODUCTION

iabetes mellitus is a major and growing public health problem throughout the world, with an estimated worldwide prevalence of 150 million people in 2000, which is expected to increase to 320 million by 2025 [1]. Besides hyperglycemic, several other factors including dislipidemia or hyperlipidemia are involved in the development of micro and macro vascular complications of diabetes which are the major causes of morbidity and death [2]. Although numerous oral hypoglycemic drugs exist alongside insulin, still there is no promising therapy to cure diabetes [3]. India has a rich emporium of various potent herbs and herbal components for treating various diseases including diabetes. In recent years, numerous traditional medicinal plants were tested for their antidiabetic potential in the experimental animals [4,5,6].

Eugenia floccosa Bedd is one of the medicinally important plants belongs to Myrtaceae family. The leaf paste of *E. floccosa* is given to treat rheumatic pain by Kanikkar tribe of Agasthiarmalai Biosphere Reserve, Tamil Nadu. The ethanol extract of *E. floccosa* has been reported for its anti-tumour activity [7]. The current investigation is an attempt to study the antidiabetic, antihyperlidaemic and antioxidant activities of ethanol extract of *E. floccosa* leaf in alloxan induced diabetic rats.

MATERIALS AND METHODS Plant Material

The leaves of *Eugenia floccosa* Bedd were freshly collected from the well grown healthy plants inhabiting the natural forests of Kothiayar, Agasthiarmalai Biosphere Reserve, Western Ghats, Tamilnadu. The plant were identified and authenticated in Botanical Survey of India, Southern Circle, Coimbatore, Tamilnadu, India. A voucher specimen was deposited in Ethnopharmacology Unit, Research Department of Botany, V.O.Chidambaram College, Tuticorin, Tamilnadu.

Preparation of plant extract for phytochemical screening and antidiabetic studies

The *E. floccosa* leaves were shade dried at room temperature and the dried leaves were powdered in a Wiley mill. Hundred grams of powdered *E. floccosa* leaves was packed in a Soxhlet apparatus and extracted with ethanol The extract were subjected to qualitative test for the identification of various phytochemical constituents as per the standard procedures [8,9,10]. The ethanol extracts were concentrated in a rotary evaporator. The concentrated ethanol extract were used for antidiabetic studies.

Animals

Normal healthy male Wistar albino rats (180- 240g) were housed under standard environmental conditions at temperature $(25\pm2^{\circ} \text{ C})$ and light and dark (12: 12 h). Rats were fed with standard pellet diet (Goldmohur brand, MS Hindustan lever Ltd., Mumbai, India) and water *ad libitum*.

Acute Toxicity Study

Acute oral toxicity study was performed as per OECD – 423 guidelines (acute toxic class method), albino rats (n=6) of either sex selected by random sampling were used for acute toxicity study [11]. The animals were kept fasting for overnight and provided only with water, after which the extracts were administered orally at 5mg/kg body weight by gastric intubations and observed for 14 days. If mortality was observed in two out of three animals, then the dose administered was assigned as toxic dose. If mortality was observed in one animal, then the same dose was repeated again to confirm the toxic dose. If mortality was not observed, the procedure was repeated for higher doses such as 50,100, and 2000 mg/kg body weight.

Induction of Experimental Diabetes

Rats were induced diabetes by the administration of simple intraperitioneal dose of alloxan monohydrate (150 mg/kg) [12]. Two days after alloxan injection, rats

^{*}Corresponding Author E-mail: vrmohan_2005@yahoo.com

screened for diabetes having glycosuria and hypoglycemia with blood glucose level of 200-260 mg/100 ml were taken for the study. All animals were allowed free access to water and pellet diet and maintained at room temperature in plastic cages.

Experimental Design

In the present investigation, a total of 30 rats (24 diabetic surviving rats and 6 normal rats) were taken and divided into five groups of 6 rats each.

- Group I: Normal untreated rats
- Group II: Diabetic control rats
- Group III: Diabetic rats given ethanol extract of *E. floccosa* leaf (150mg/kg body weight)
- Group IV: Diabetic rats given ethanol extract of *E. floccosa* leaf (300mg/kg body weight)
- Group V: Diabetic rats given standard drug glibenclamide (600mg/kg body weight).

Biochemical analysis

The animals were sacrificed at the end of experimental period of 14 days by decapitation. Blood was collected, sera separated by centrifugation at 3000g for 10 minutes. Serum glucose was measured by the O-toluidine method [13]. Insulin level was assayed by Enzyme Linked Immunosorbant Assay (ELISA) kit [14]. Urea estimation was carried out by the method of Varley [15]; serum creatinine was estimated by the method of Owen et al [16]. Glycosylated haemoglobin (HBA₁C) estimation was carried out by a modified colorimetric method of Karunanayake and Chandrasekharan [17]. Serum total cholesterol (TC) [18], total triglycerides (TG) [19], low density lipoprotein cholesterol (LDL-C), very low density lipoprotein cholesterol (VLDL- C) [20], high density lipoprotein cholesterol (HDL-C) [21] and phospholipids [22] were analyzed. Serum protein [23] and serum albumins were determined by quantitative colorimetrically method by using bromocresol green. The total protein minus the albumin gives the globulin, serum glutamate pyruvate transaminase (SGPT) and serum glutamate oxaloacetate transaminase (SGOT) were measured spectrophotometrically by utilizing the method of Reitman and Frankel [24]. Serum alkaline phosphatase (ALP) was measured by the method of King and Armstrong [25]. Catalase (CAT) [26], superoxide dismutase (SOD) [27], lipid peroxidation (LPO) [28], reduced glutathione (GSH) [29] and glutathione peroxidase (GPx) [30] were analyzed in the normal, diabetic induced and drug treated rats.

STATISTICAL ANALYSIS

The data were analyzed using student's t-test statistical methods. For the statistical tests a p values of less than 0.01 and 0.05 was taken as significant.

RESULTS AND DISCUSSION

The phytochemical screening of ethanol extract of E. floccosa leaf revealed the presence of alkaloid, catechin, coumarin, tannin, saponin, steroid, flavonoid, phenol, sugar, glycoside, xanthoprotein and fixed oil. Acute toxicity study revealed the non-toxic nature of the ethanol extract of E. floccosa leaf. Table 1 shows the levels of blood glucose, plasma insulin, urea, creatinine and glycosylated haemoglobin of normal, diabetic rats and drug treated rats. The alloxan induced diabetic rats elicited significant rise in blood glucose from 69.50 to 201.00mg/dl (p<0.05) and a significant decrease in plasma insulin level from 24.50 to 5.40 (p<0.01). On the contrary, diabetic rats treated with ethanol extract of E. floccosa leaf exhibited decrease in blood glucose and increase in plasma insulin significantly at a dose of 150 mg/kg and 300 mg/kg body weight. It was observed that ethanol extract of E. floccosa reversed these effects in diabetic animals. The possible mechanism by which ethanol extract brings about its hyperglycemic action may be by induction of pancreatic insulin secretion from β cells of islets of langerhans or due to enhanced transport of blood glucose to peripheral tissue [31]. Earlier many plants have been studied for their hypoglycemic and insulin release stimulatory effects [32,33,34,35,6,36].

A significant elevation in serum constituents, urea and creatinine were observed in alloxan induced diabetic rats (Group II), when compared to control rats. The ethanol extracts of E. floccosa leaves were administered orally (150 mg/ kg body weight- Group III and 300mg/kg body weight- Group IV) to rats for 14 days, reversed the urea and creatinine level to near normal. The administration of glibenclamide (Group V) also decreased the levels of urea and creatinine to some extent. Alloxan is taken as indications of an abnormal glomerular fraction where a simple injection of cisplation at a dose of 5mg/kg body weight in rabbits caused a marked reduction in the glomerular filtration rates, which was accompanied by an increase in the creatinine level, indicating the induction of acute renal failure. It is confirmed that there is a significant increase in serum creatinine in albino rats 14 days after alloxan administration. The present results show that, the treatment with ethanol extract of E. floccosa leaf was effective in preventing alloxan induced increase in serum creatinine level when compared with the control. Alloxan induced diabetic rats showed significant increase (p < 0.05) glycosylated haemoglobin (HBA1C) level compared with normal rats. The ethanol extract of E. floccosa leaf treated rats showed a significant decrease (p<0.05) in the content of glycosylated haemoglobin. Glycosylated haemoglobin determinations are self monitoring of blood glucose therefore play an important complementary roles for the management of diabetes mellitus [37].

The levels of serum protein, albumin and globulin of control, alloxan induced diabetic rats and drug treated rats were presented in Table 2. A significant reduction in serum protein, albumin and globulin were observed in alloxan induced

Parameter	Insulin (Mlu/ml)	Glucose (mg/dl)	Urea (mg/dl)	Creatinine (mg/dl)	Glycosylated Hb (%)
Group I	24.50±1.4	69.50±1.2	11.54±1.9	0.61±0.8	3.6±0.05
Group II	05.40±0.6**	201.00±11.2*	39.51±5.6*	1.72±0.7	12.51±1.1*
Group III	12.60±1.1*a	105.50±6.3*a	26.21±4.2	1.23±0.1	9.36±1.7*
Group IV	17.80±1.3** a	76.50±57aa	9.34±5.1aa	0.89±0.4	7.07±1.03a
Group V	22.50±0.8*	91.50±6.9aa	12.74±1.9a	0.81±0.5	4.91±0.7

Table 1: Effect of ethanol extracts of *Eugenia floccosa* leaves on the serum insulin, glucose, urea, creatinine and HBA¹C level of normal, diabetic induced and drug treated adult albino rats.

Each value is SEM of 6 animals. Comparison made between normal control to diabetic control and drug treated groups * p < 0.05; **p<0.01 and comparison made between diabetic control to drug treated groups a p<0.05; aa p<0.01

Table 2: Effect of ethanol extract of *Eugenia floccosa* leaves on the serum protein, albumin, globulin, SGOT, SGPT and ALP level of normal, diabetic induced and drug treated adult albino rats.

Parameter	Protein (g/dl)	Albumin (g/dl)	Globulin(g/dl)	SGPT (u/l)	SGOT (u/l)	ALP (u/l)
Group I	8.94±0.11	4.14±0.64	4.87±0.11	19.2±3.2	21.4±3.3	164.55±5.4
Group II	6.51±0.71*	3.91±0.35	2.65±0.08*	35.4±6.2*	35.3±5.9*	196.65±3.4
Group III	7.14±0.92	4.04±0.87	3.17±0.10	29.1±3.2	27.4±4.1	113.42±4.4
Group IV	8.14±0.89	4.99±0.43	4.15±0.06	20.6±3.4	24.5±2.8	151.43±5.6
Group V	7.94±0.30*	4.11±0.32	3.83±0.04	16.5±4.8*	28.4±2.2	147.33±5.7

Each value is SEM of 6 animals. Comparison made between normal control to diabetic control and drug treated groups * p < 0.05

Table 3: Effect of ethanol extract of *Eugenia floccosa* leaves on the serum lipid profile of normal, diabetic induced and drug treated adult albino rats.

Parameter	TC (mg/dl)	TG(mg/dl)	LDL-C (mg/dl)	VLDL (mg/dl)	HDL (mg/dl)	PL (mg/dl)
Group I	104.51±3.56	88.51±2.84	52.59±2.05	17.70±1.22	34.21±0.52	161.01±13.91
Group II	184.53±8.42*	206.55±8.41***	116.69±6.25*	41.31±3.45*	26.53±3.84*	232.23±12.17*
Group III	121.51±6.31	151.56±3.99**	60.38±2.33a	30.31±1.23*	30.81±1.91	176.14±16.39
Group IV	116.64±5.21	109.32±4.14*a	55.44±5.55a	21.86±2.33**a	32.33±2.11	171.80±10.96
Group V	113.56±4.52	109.53±7.3*a	62.34±4.12	21.90±1.98*a	29.31 ±4.23	169.06±13.84aa

Each value is SEM of 6 animals. Comparison made between normal control to diabetic control and drug treated groups * p < 0.05; **p < 0.01; *** p < 0.001 and comparison made between diabetic control to drug treated groups a p < 0.05; as p < 0.01

diabetic rats (Group II) when compared to control (Group I) and glibenclamide treated rats (Group V). On administration of ethanol extract of *E. floccosa* to the diabetic rats, the levels of protein, albumin and globulin were found to be restored in normal. These results were in accordance with the effects of *Wattakaka volubilis* [6], *Senna auriculata* [38] and *Pterocarpus marsupium* [36] in diabetic rats.

Table 2 summarized the effect of alloxan on the activity of the hepatic marker enzymes in serum. In the present study, the levels of SGPT and SGOT in alloxan induced diabetic rats were elevated. It may be due to leaking out of enzymes from the tissues and migrating into the circulation by the adverse effect of alloxan [34]. Aspartate amino transaminases and Alanine transaminase were used as markers to assess the extent of liver damage in streptozotocin induced diabetic rats [39]. In this study, the ethanol extract of *E. floccosa* regulated the activity of SGPT and SGOT in liver of rats intoxicated with alloxan. The effect of glibenclamide on the recovery of hepatic enzyme activity in serum was very similar to that of the earlier study [40].

The restorations of SGPT and SGOT to their respective normal levels after treatment with both glibenclamide and ethanol extract of *E. floccosa*, further strengthen the antidiabegenic effect of this extract. Moreover, SGPT and SGOT levels also act as indicators of liver function and restoration of normal levels of these parameters indicate normal functioning of liver. Since the alloxan can also affect the liver by free radical mechanism.

In addition to the assessment of SGPT and SGOT levels during diabetes the measurement of enzymatic activities of phosphatases such as acid phosphatase (ACP) and alkaline phosphatase (ALP) is of clinical and toxicological importance as changes in their activities are indicative of tissue damage by toxicants. In the present study, serum ALP increased in alloxan induced diabetic rats (Table 2). Elevated level of this enzyme in diabetes may be due to extensive damage to liver in the experimental animal by alloxan. Treatment with ethanol extract of *E. floccosa* in alloxan induced diabetic rats produces a decline in ALP level.

The levels of serum lipid profile, total cholesterol (TC), triglycerides (TG), LDL-C, VLDL-C and HDL-C in control, diabetic induced and drug treated rats were presented in Table 3. Alloxan induced rats showed significant increase in serum lipid profiles except HDL-C when compared with normal rats. The glibenclamide (Group V) and ethanol extract of *E. floccosa* (Group III and IV) treated rats

Parameter	LPO (mmole/dl)	SOD (unit x/mg protein)	CAT (unit y/mg protein)	GPX(unit z/mg protein)	GSH (mg/dl)
Group I	0.83± 0.34	6.93± 0.74	0.49± 0.31	7.36± 0.81	91.54± 3.05
Group II	3.11± 0.98*	14.56± 0.54**	2.96± 0.88*	13.23± 0.92**	27.31± 2.86**
Group III	1.15 ±0.88	9.39± 0.65	0.57± 0.77a	8.59 ±1.65	42.16±1.54a
Group IV	0.74 ± 0.41a	4.91 ± 0.33a	0.45 ± 3.9a	5.81 ± 1.64a	89.62±3.85
Group V	0.89± 0.14a	7.99± 0.33	0.84± 0.14	8.03± 0.65a	87.4± 5.18a

Table 4: Effect of ethanol extracts of *Eugenia floccosa* leaves on the LPO, SOD, CAT, GPX and GSH enzymes in the plasma of normal, diabetic induced, and drug treated adult albino rats.

Each value is SEM of 6 animals. Comparison made between normal control to diabetic control and drug treated groups. * p < 0.05; **p < 0.01 and comparison made between diabetic control to drug treated groups a p < 0.05. x-One unit of SOD is defined as the enzyme concentration which gives 50% inhibition of NBT reduction in one minute. y-One unit of CAT is defined as the μ mole of hydrogen peroxide consumed per minute. z- One unit of GPx is defined as the μ g of glutathione consumed per minute.

showed a significant decrease in the content of lipid profile when compared with diabetic induced rats. Similarly HDL-C level decreased in alloxan induced diabetic rats when compared to normal rats. On administration of ethanol extract of E. floccosa and glibenclamide to the diabetic rats, HDL-C level was found to be restored to normal. The level of serum lipid profiles are usually raised in diabetic rats in the present study and such elevation represents risk factor for coronary heart diseases [41]. Lowering of the serum lipid level through dietary or drug therapy seems to be associated with a decrease in the risk of vascular disease [42]. During diabetes, enhanced activity of the enzyme, increased lipolysis and releases more fatty acids into the circulation [43]. The increased fatty acid concentration also increases the β -oxidation of fatty acids, producing more acetyl Co-A and cholesterol during diabetes. In normal condition, insulin increases receptor-mediator removal of LDL-cholesterol and decreased activity of insulin, during diabetes causes hypercholesterolemia. Hypercholesterolemia and hypertriglycedemia have been reported to occur in diabetic rats [41]. The increased concentration of free fatty acid may be due to lipid break-down and this may cause increased generation of NADPHdependent microsomal lipid peroxidation. Phospholipids were increased in alloxan induced diabetic rats. Phospholipids are present in cell membrane and make up vast majority of the surface lipoprotein forming a lipid bilayer that acts as an interface with both polar plasma environment and non-polar lipoprotein of lipoprotein core [44]. Increased phospholipids levels in tissues were reported by Venkateswaran et al. [45]; Pari and Satheesh, [46] in streptozotocin diabetic rats. Administration of ethanolic extract of E. floccosa leaf and glibenclamide decreased the levels of phospholipids.

The results (Table 4) showed increased lipid peroxidation (LPO) of alloxan induced diabetic rats. Earlier studies have reported that there was an increased lipid peroxidation in liver, kidney and brain of diabetic rats [47,48]. This may be because the tissues contain relatively high concentration of early peroxidizable fatty acids. In the present study, an increase in the levels of LPO was found and these levels were significantly reduced after the supplementation of the ethanol extract of *E. flocossa* leaf and glibenclamide (Table 4). This indicate that plant extract inhibit oxidative damage due to the antiperoxidative effect of ingredients present in ethanol extract of *E. flocossa*. This could be correlated with previous study reported that *Cassia auriculata* flower [35] *Syzigium cuminii* [49,50] *Tinospora cardifolia* [51] and *Scoparia dulcis* [52] has antiperoxidative and antihyperlipidemic effect of diabetic animals. Apart from the regulation of carbohydrate metabolism, insulin also plays an important role in the lipid metabolism. Insulin is a potent inhibitor of lipolysis, since it inhibits the activity of hormone sensitive lipase in adipose tissue and suppresses the release of free fatty acids [53].

The levels of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and reduced glutathione (GSH) (Table 4) were significantly

(p<0.05) reduced in alloxan induced rats. These adverse changes were reversed to near normal values in ethanol extract of E. floccosa leaf treated. It is well known that CAT, SOD and GPx play an important role as protective enzymes against free radical formation in tissues [54]. It has been shown that the antioxidants status of tissues is an important factor in the development of diabetic complications [55]. Reduced glutathione plays an important role mainly in the detoxification and metabolism as a cofactor or a substrate for some enzymes and in this way it is an antioxidant agent protecting tissues from oxidative stress and thus measured as a common marker of free radical damage. Enzymatic antioxidant such as SOD and CAT are considered primary enzymes since they are involved in the direct elimination of ROS [56] SOD is an important defense enzyme and scavengers O2 anion from H2O2 and hence diminishes the toxic effects due to this radical or other free radicals derived from secondary reaction [57]. CAT is a haemoprotein, which catalyzes the reduction of hydrogen peroxides [58]. The antioxidant enzymes such as SOD and CAT are known to be inhibited in diabetes mellitus as a result of non- enzymatic glycosylation and oxidation [59]. In the present study, the activities of SOD and CAT decreased in diabetic rats as reported earlier, which could be due to inactivation caused by alloxan generated ROS [60]. The ethanol extract of E. floccosa had reversed the activities of these enzymatic antioxidants, which might be due to decreased oxidative stress as evidenced by decreased LPO.

In conclusion, the present study has shown that the ethanol extract of *E. floccosa* leaf have antidiabetic, antihyperlipidaemic and antioxidant effects. The possible antidiabetic activity of the extracts might be due to stimulation of residual pancreatic insulin or by increasing peripheral utilization of glucose. Glycosides, flavonoids, tannins, organic sulphur compounds, catechol and alkaloids are active ingredients of hypoglycemic plant [61]. Flavonoids are reported to regenerate the damaged pancreatic beta cells [62]. Phenols have found to be effective antihyperglycemic agents [63]. In the present study, the phytochemical analysis of ethanol extract of *E. floccosa* clearly pointed out the presence of above said active phytochemicals. It denotes that the antidiabetic effect of ethanol extract of *E. floccosa* may be due to the presence of more than one antihyperglycemic principle and their synergistic effects.

ACKNOWLEDGEMENT

Thanks to Dr. Sampathraj, Honorary Advisor, Samsun Clinical Research Laboratory, Tirupur for their assistance in animal studies.

REFERENCES

 Sicree R, Shaw J, Zimmet P. Diabetes and impaired glucose tolerance In: Gan. D (Ed.) Diabetes Atlas. International Diabetes Federation, 3rd edn. International diabetes Federation. Brussels, Belgium 2006; pp15-103.

- Bennet PH, Joshins. Diabetes Mellitus Kahan R.C and Weir (Eds.) Baltimore international Publication. Waverley 1998; pp: 193-200.
- Sumana G, Suryawarshi SA. Effect of Vinca rosea extracts in treatment of alloxan diabetes in male albino rats. International Journal of Experimental Biology. 2001; 39: 748-758.
- Srivastava Y, Bhat HV, Verma Y, et al. Antidiabetic and adaptogenic properties of *Momodica charantia* extract: An experimental and clinical evaluation. Phytotherapy Research. 1993; 7: 285.
- Bhaskar VH, Ajay S. Antihyperglycemic and antihyperlipidaemic activities of root extracts of *Calatropis procera* (Ait) R.Br. on streptozotocin induced diabetic rats. Jordan Journal of Biological Sciences. 2009; 2: 177-180.
- Maruthupandian A, Mohan VR, Sampathraj R. Antidiabetic, antihyperlipidaemic and antioxidant activity of *Wattakaka volubilis* (L.f) Stapf leaves in alloxan induced diabetic rats. International Journal of Pharmaceutical Science and Research. 2010; 1: 83-90.
- Kala SMJ, Tresina Soris P, Mohan VR. Antitumour activity of Eugenia floccosa Bedd and Eugenia singampattiana Bedd leaves against Dalton ascites lymphoma in swiss albino mice. International Journal of PharmTech Research. 2011; 3: 1796-1800.
- Brinda P, Sasikala P, Purushothaman KK. Pharmacognostic studies on Merugan kizhangu. Bulletin of Medicinal and Ethnobotanical Research. 1981; 3: 84-96.
- Anonymous. Phytochemical investigation of certain medicinal plants used in Ayurveda. Central Council for Research in Ayurveda and Siddha, New Delhi. 1990; 1-356.
- 10. Lala PK. Lab manuals of Pharmacognosy CSI Publishers and Distributers, Kolkata. 1993.
- OECD. Organisation for Economic co-operation and Development). OECD guidelines for the testing of chemicals/Section 4: Health Effects Test No. 423; Acute oral Toxicity- Acute Toxic Class method. OECD. Paris. 2002.
- Nagappa AN, Thakurdesai PA, Venkat Rao N, et al. Antidiabetic activity of *Terminalia catappa* Linn. fruits. Journal of Ethnopharmacology 2003; 88: 45-50.
- Sasaki T, Masty S, Sonae A. Effect of acetic acid concentration on the colour reaction in the Otoluidine boric acid method for blood glucose estimation. *Rinshbo Kagaku.*, 1992; 1: 346-353.
- Anderson L, Dinesen B, Jorgonsen PN, et al. Enzyme immune assay for intact human insulin in serum or plasma. Clinical Chemistry. 1993; 39: 578-582.
- 15. Varley H. Practical clinical biochemistry, Arnold Heinemann Publication Pvt. Ltd., 1976; p, 452.
- Owen JA, Iggo JB, Scangrett FJ, et al. Determination of creatinine in plasma serum, a critical examination. Journal of Biochemistry. 1954; 58: 426-437.
- Karunanayake EH , Chandrasekharan NV. An evaluation of a colorimetric procedure for the estimation of glycosylated haemoglobin and establishment of reference values for Sri Lanka. Journal of Natural Science Council of Sri Lanka. 1985; 13: 235-258.
- Parekh AC, Jung. Cholesterol determination with ferric acetate, uranium acetate and sulphuric acid, ferrous sulphate reagent. Analytical Chemistry. 1970; 112: 1423-1427.
- Rice EW. Triglycerides in Serum In: Standard Methods. Clinical Chemistry. 9ed Roderick MP, Academic press, New York. 1970; 215-222.
- Friedwald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low density lipoprotein cholesterol in plasma, without use of the preparative ultra centrifuge. Clinical Chemistry.1972; 18: 499-502.
- Warnick GR, Nguyan T, Albers AA. Comparison of improved precipitation methods for quantification of high density lipoprotein cholesterol. Clinical Chemistry. 1985; 31: 217.
- Takayama M, Itoh S, Nagasaki T, et al. A new enzymatic method for determination of serum phospholipids. Clinical Chemistry Acta. 1977; 79: 93 – 98.
- 23. Lowry OH, Rosenbrough NJ, Farr AL et al. Protein measurement with the folin's phenol reagent. Journal of Biological Chemistry. 1951; 193: 265-275.

- Reitman S, Frankel SA. Colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. American Journal of Clinical Pathology. 1957; 28: 56-63.
- 25. King EJ, Armstrong AR. Determination of serum and bile phosphatase activity. Cannadian Medical Association Journal. 1934; 31: 56-63.
- Bergmayer HU. UV method of catalase assay. In Methods of Enzymatic Analysis, Weiheim Deer field Beach, Florida, Bansal. 1983; 3: 273.
- Madesh M , Balasubramanian KA. Microtitre plate assay for superoxide dismutase using MTT reduction by superoxide. 'Indian Journal of Biochemistry and Biophysics. 1998; 35: 184-188.
- Rehman S. Lead induced regional lipid peroxidation in brain. Toxicological Letters. 1984; 21: 333-337.
- Prins HK, Loos JA. In Glutathione; Biochemical methods in red cell genetics, edited by J.J Yunis. Academic Press, New York. 1969; 127-129.
- Pagila DE, Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. Journal of Laboratory and Clinical Medicine. 1967; 70: 158-169.
- Hakkim FL, Girija S, Senthilkumar R et al. Effect of aqueous and ethanol extracts of *Cassia auriculata* L. flowers on diabetic using alloxan induced diabetic rats. International Journal of Diabetic Metabolism. 2007; 15: 100-106.
- Morrison EY, Smith SA, West M, et al. The effect of *Bixa orellana* on blood sugar levels in the anaesthetized dog. West Indian Medical Journal. 1985; 34: 38-42.
- Al-Hader AA, Haesan ZA, Agel MB. Hyperglycemic and insulin release inhibitory effects of *Rosmarinus officinalis*. Journal of Ethnopharmacology. 1994; 36: 99-103.
- Stanely P, Prince M, Menon V. Hypoglycemic and other related actions of *Tinospora cordifolia* roots in alloxan induced diabetic rats. Journal of Ethnopharmacology. 1999; 70: 9 – 15.
- Pari L, Latha M. Effect of Cassia auriculata flowers on blood sugar levels, serum and tissue lipids in Streptozotocin Diabetic Rats. Singapore Medicinal Journal. 2002; 43: 617-621.
- Maruthupandian A, Mohan VR. Antidiabetic, antihyperlipidaemic and antioxidant activity of *Pterocarpus marsupium* Roxb. in alloxan induced diabetic rats. International Journal of PharmTech Research. 2011; 3: 1681-1687.
- Thai AC, Yeo PPB, Chan L, et al. Glycosylated haemoglobin and diabetic control. Singapore Medicinal Journal. 1983; 24: 210-212.
- Shanmugasundaram R, Kalpana Devi V, Tresina Soris P, et al. Antidiabetic, antihyperlipidaemic and antioxidant activity of *Senna auriculata* (L.) Roxb leaves in alloxan induced diabetic rats. International Journal of PharmTech Research. 2011; 3: 747-756.
- Hwang HJ, Kim SW, Lim JM, et al. Hypoglycemic effect of crude epoxypolysaccharides produced by a medicinal mushroom *Phellinus baumii* in streptozotocin induced diabetic rats. Life Science. 2005; 76: 3069 – 3080.
- Preethi KC, Kuttan R. Hepato and reno protective action of *Calendula* officinalis L. flower extract. Indian Journal of Experimental Biology. 42009; 7: 163 – 168.
- 41. Mironova MA, Klein RL, Virella GL, et al. Anti-modified LDL antibodies, LD -containing immune complexes and susceptibility of LDL to *in vitro* oxidation in patients with type 2 diabetes. Diabetes. 2000; 49: 1033-1049.
- 42. Scott M, Grundy MD. Diabetic cardiovascular Disease. Circulation. 1999; 100: 1134-1146.
- Agardh CD, Bjorgell P, Nilson EP. The effect of tolbutamide on lipoproteins and lipoproteinlipase and hormone sensitive lipase. Diabetes Research Clinical Practice 1999; 46: 99-108.
- Cohn RM, Roth KS. Lipid and lipoprotein metabolism In: Biochemistry and Diseases, Williams and Willkins publishers, Baltimore, 1996; p 280.
- Venkateswaran S, Pari L, Saravanan G. Effect of *Phaseolus vulgaris* on circulatory antioxidants and lipids in streptozotocin-induced diabetic rats. Journal of Medicinal Food. 2002; 5: 97 – 104.
- Pari L, Satheesh AM. Antidiabetic effect of *Boerhavia diffusa*: effect on serum and tissue lipids in experimental diabetes. Journal of Medicinal Food. 2004; 7: 472 476.
- Latha M, Pari L Preventive effects of Cassia auriculata L. flowers on brain lipid peroxidation in rats treated with streptozotocin. Molecular Cell and Biochemistry. 2003a; 243: 23-28.

- Ananthan R, Latha M, Ramkumar KM, et al. Antidiabetic effect of Gymnema montanum leaves: effect on lipid peroxidation induced oxidative stress in experimental diabetes. Nutrition. 2004; 6: 379 – 386
- Prince PSM, Menon VP. Hypoglycemic activity of Syzigium cumini seeds; effect on lipid peroxidation in alloxan diabetic rats. Journal of Ethnopharmacology 61, 1998: 1 – 7.
- Prince PSM, Kamalakkannan N, Menon VP. Antidiabetic and antihyperlipidaemic effect of alcoholic Syzigium cumini seeds in alloxan induced diabetic albino rats. Journal of Ethnopharmacology 91, 2004: 209 – 213.
- Prince PSM, Menon VP, Gunasekaran G. Hypolipidemic action on *Tinospora cordifolia* root extract in alloxan diabetic rats. Journal of Ethnophamacology 14, 1999: 4 – 16.
- Latha M, Pari L. Modulatory effect of *Scoparia dulcis* in oxidative stress induced lipidperoxidation in streptozotocin diabetic rats. Journal of Medicinal Food. 2003b ; 6: 379 – 386.
- 53. Loci AS, Shaabha M, Khazraji AL, et al. Hypoglycemic effect of a valuable extract of *Artemisia herba-alba* II. Effect of a valuable extract on some blood parameters in diabetic animals. Journal of Ethnopharmacology. 1994; 43: 167 - 171.
- 54. Oberly WR and Buettner RG. Role of superoxide dismutase in cancer. Cancer Research. 1974; 35: 1141-1149
- Wohaeib SA, Godin DV. Alterations in free radical tissue defense mechanisms in streptozotocin-induced diabetic rats. Effect of insulin treatment. Diabetes. 1987; 6: 1014-1018.

- 56. Arulselvan P, Subramanian SP. Beneficial effects of *Murraya koenijii* leaves on antioxidants defense system and ultra structural changes of pancreatic cells in experimental diabetes in rats. Chemico-Biological Interactions. 2007; 165: 155-164.
- Manonmani G, Bhavapriya V, Kalpana S, et al. Antioxidant activity of Cassia fistula Linn. Flowers in alloxan induced diabetics rats. Journal of Ethnopharmacology. 2005; 97: 39-42.
- Punitha ISR, Shirwaikar A, Shirwaikar A. Antidiabetic activity of benzyl tetra isoquinoline alkaloid berberine in streptozotocin- nicotinamide induced type 2 diabetic rats. Diabetologia Croatica. 2005; 34: 177-128.
- Al-Azzaawie H, Alhamdani MSS. Hypoglycemic and antioxidant effect of oleuropein in alloxan- diabetic rabbits. Life Science. 2006; 78: 1371-1377.
- Sepici A, Gurbuz I, Cevik C, et al. Hypoglycemic effects of myrtle oil in normal and alloxan- diabetic rabbits. Journal of Ethnopharmacology. 2004; 93: 311-318.
- Oliver B. Oral hypoglycaemic plants in West Africa. Journal of Ethnopharmacology 1980; 2: 119-127.
- Chakravarthy BK, Gupta S, Gambir SS et al. Pancreatic beta cell regeneration. A novel antidiabetic mechanism of *Pterocarpus marsupium* Roxb. Indian Journal of Pharmacology. 1980; 12: 123 – 127.
- Manickam M, Ramanathan M, Farboodinary Jahromi MA, et al. Antihyperglycemic activity of phenolic form *Pterocarpus marsupium*. Journal of Natural Product. 1997; 60: 609-610.