

Journal of Basic and Clinical Pharmacy

www.jbclinpharm.com

entricitival JBCP

An Antioxidant potential of Hingot and Hadjod

Priyanka Vijay^{*1} and R. Vijayvergia¹

1) Plant Pathology and Biochemistry Laboratory, Department of Botany, University of Rajasthan Jaipur, 302004.

ABSTRACT: The antioxidant activity of two plants - Hadjod i.e. *Cissus quadrangularis* (CQ) and Hingot i.e. *Balanites aegyptiaca* (BA) was determined by the thiocyanate method. The antioxidant activity of both the plants increased with increasing amount of extract (200 g-1000 g) added to the linoleic emulsion. The ethanolic extract of CQ was more effective than the other. Like antioxidant activity, the reducing power was also dependent upon the concentration. The ethanolic extract of BA shows more reducing power than the other. The result obtained in the present study indicates that the both the plants are potential source of natural antioxidants. In addition, we could suggest that although the reducing power of a substance may be an indicator of its potential antioxidant activity, there is not necessarily a linear correlation between these two activities.

KEY WORDS:

Cissus quadrangularis Balanites aegyptiaca linoleic emulsion antioxidant, reducing power received on 05-01-2010 modefied on 15-01-2010 accepted on 24-01-2010 available online 15-02-2010 www.jbclinpharm.com

INTRODUCTION:

There is an increased evidence for the participation of free radicals in the etiology of various diseases like cancer, diabetes, cardiovascular diseases, autoimmune disorders, neurodegenerative diseases, aging, etc. ¹. A free radical is defined as any atom or molecule possessing unpaired electrons. The primary oxygen derived free radicals are superoxide anion (O2.), hydroxyl (OH.), hydroperoxyl (OOH-), peroxyl (ROO-) and alkoxyl (RO.) radicals and non free radicals are hydrogen peroxide (H2O2), hypochlorous acid (HOCl), ozone (O3) and singlet oxygen (1O2). These reactive intermediates are collectively termed as reactive oxygen species (ROS). Free radicals can cause a wide range of toxic oxidative reactions like initiation of the peroxidation of the membrane lipids leading to the accumulation of lipid peroxides, direct inhibition of mitochondrial respiratory chain enzymes, fragmentation or random cross linking of molecules like DNA, enzymes and proteins which ultimately leads to cell death². ROS can be formed in living organisms by both endogenous and exogenous sources. Endogenous sources of free radicals include normal

aerobic respiration, peroxisomes and stimulation of polymorphonuclear leucocytes and macrophages. The exogenous sources include ionizing radiation, tobacco smoke, pollutants, pesticides and organic solvents³.

Antioxidants are agents which scavenge the free radicals and prevent the damage caused by them. They can greatly reduce the damage due to oxidants by neutralizing the free radicals before they can attack the cells and prevent damage to lipids, proteins, enzymes, carbohydrates and DNA⁴. Antioxidants can be classified into two major classes i.e., enzymatic and non-enzymatic. The enzymatic antioxidants are produced endogenously and include superoxide dismutase, catalase, and glutathione peroxidase. The non-enzymatic antioxidants include tocopherols, carotenoids, ascorbic acid, flavonoids and tannins which are obtained from natural plant sources⁵. A wide range of antioxidants from both natural and synthetic origin has been proposed for use in the treatment of various human diseases⁶. There are some synthetic antioxidant compounds such as butylated hydroxytoluene, butylated hydroxyanisole and tertiary butylhydroquinone which are commonly used in processed foods. However, it has been suggested that these compounds have shown toxic effects like liver damage and mutagenesis^{7,8}. Flavonoids and other

^{*}Corresponding Author:

E-mail: jaaldhi@sify.com

phenolic compounds of plant origin have been reported as scavengers of free radicals^{9,10}. Hence, nowadays search for natural antioxidant source is gaining much importance. Therefore, attempt has been made to evaluate antioxidant potential of selected Indian medicinal plants viz-, *Balanites aegyptiaca and Cissus quadrangularis*.

Balanites aegyptiaca (Zygophyllaceae) commonly known as 'Desert Palm' is widely grown in African continents, middle-east, south Asia. The plant is also known as 'Pharmacy tree' because all parts of the plant are used for medicinal purposes¹¹. Chapaganin *et al.*, (2005)¹² observed that the plant is used for human consumption and cosmectics. *Cissus quadrangularis* (Vitaceae) commonly known as Hadjod is used in Ayurveda in Calciferous process, in back, spine, bones.^{13,14}

MATERIAL AND METHODS

Plant material

The plant materials were collected in August 2007 from the adjacent areas of Jaipur district, and were taxonomically identified by Roop Singh ji, a plant taxonomist. A voucher specimen (*Balanites aegyptiaca*-RUBL20252, *Cissus quadrangularis*-RUBL20253) has been preserved in the departmental herbarium, Department of Botany, University of Rajasthan, Jaipur, India.

Preparation of extract

The air-dried powdered leaves of *B.aegyptiaca*, *C. quadrangularis* (100 g) were extracted with 450 ml of ethanol mixture using a mechanical shaker for 4 h. The resultant extract was concentrated under reduced pressure to yield a residue

Antioxidant activity

The peroxy radical scavenging activity was determined by thiocyanate method using ascorbic acid (50-800 μ g/ml) as standard(15). Increasing concentration of the fractions (50-800 μ g/ml) in 0.5 ml of distilled water was mixed with 2.5 ml of 0.02 M linoleic acid emulsion (in 0.04 M phosphate buffer pH 7.0) and 2 ml phosphate buffer (0.04M, pH 7) in a test tube and incubated in darkness at 37oC. At intervals during incubation, the amount of peroxide formed was determined by reading the absorbance of the red colour developed at 500 nm by the addition of 0.1 ml of 30% ammonium thiocyanate solution and 0.1 ml of 20 mM ferrous chloride in 3.5% hydrochloric acid to the reaction mixture. The percentage scavenging activity was calculated and was compared with the standard, ascorbic acid.

Reducing Power

The reducing power was investigated by the Fe³⁺- Fe^{2+} transformation in the presence of the fractions as described by Fejes et al., $(2000)^{16}$. The Fe²⁺ can be monitored by measuring the formation of Perl's Prussian blue at 700 nm¹⁷. One ml of the fraction (50-800 µg/ml), 2.5 ml of phosphate buffer (pH 6.6) and 2.5 ml of 1% potassium ferricyanide were incubated at 50°C for 30 min and 2.5 ml of 10% trichloroacetic acid was added to the mixture and centrifuged for 10 min at 3000g. About 2.5 ml of the supernatant was diluted with 2.5 ml of water and shaken with 0.5 ml of freshly prepared 0.1% ferric chloride. The absorbance was measured at 700 nm. Ascorbic acid (50-800 μ g/ml) was used as the standard. All tests were performed in triplicate and the graph was plotted with the average of the three determinations.

RESULT AND DISCUSSION

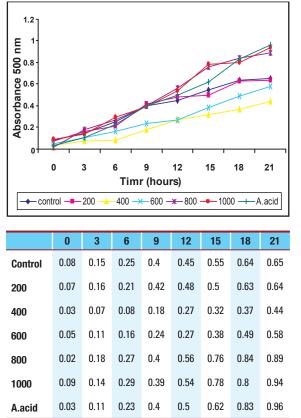
Free radicals are known to play a definite role in a wide variety of pathological manifestations. Antioxidants fight free radicals and protect us from various diseases. They exert their action either by scavenging the reactive oxygen species or protecting the antioxidant defence mechanisms.

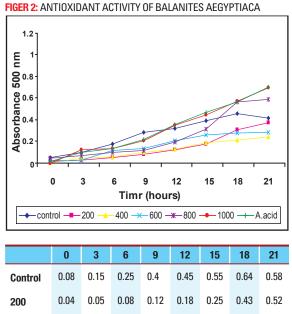
The amount of formed peroxides was measured by the thiocyanate method. The fractions were incubated with linoleic emulsion in dark at 37°C and the amount of peroxides was determined spectro-photometrically by measuring the absorbance at 500 nm¹⁹. A decrease in absorbance indicated the antioxidant activity of the fractions which might be due to the inactivation of the free radicals and the presence of flavonoid like phytochemicals.

Antioxidant activity of Hadjod (Cissus quadrangularis)

As seen in fig.1 there is no clear difference of antioxidant activity between the control and the sample containing 200 μ g extract. However peroxy radical scavenging activity is suppressed about 6h in the presence of 400 μ g extract, and after that it begins to increase. In the presence of 1000 μ g extract or 500 μ g ascorbic acid, the peroxidation process is delayed for about 12h.







0.02

0.03

0.07

0.01

0.03

400

600

800

1000

Asc A

0.07

0.05

0.11

0.18 0.2

0.15 0.2

0.09

0.17

0.15

0.15

0.2

0.17

0.29

0.31

0.19

0.29

0.28

0.49

0.5

0.27

0.37

0.44

0.62

0.65

0.3

0.39

0.78

0.79

0.78

0.34

0.4

0.82

0.96

0.97

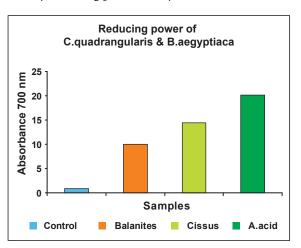
Antioxidant activity of Hingot (Balanites aegyptiaca)

As seen fig 2, the peroxy radical scavenging activity is suppressed about 6h in the presence of $400\mu g$ extract, and after that it begins to increase. In the presence of $1000\mu g$ extract or $500\mu g$ ascorbic acid, the peroxidation process is delayed for about 12h.it shows similar pattern of activity but possess less as compare to Hadjod.

Unlike antioxidant activity, the reducing power of *Cissus quadrangularis* was lowest as compare to the other one i.e. *Balanites aegyptiaca*. Although. Ascorbic acid was also more effective than the two.

It was interesting to find that although the ethanolic extract of *Cissus quadrangularis* had highest antioxidant activity, but less effective in reducing power. The reducing capacity of a compound may serve as a significant indicator of its potential antioxidant activity (Meir, 1995). However, the antioxidant activities of putative antioxidants have been attributed to various mechanisms, among which are prevention of chain initiation, binding of transition metal ion catalyst, decomposition of peroxides, prevention of abstraction, and radical scavenging²⁰. Hence, we can suggest that there is always no linear correlation between total antioxidant activity and reducing power activity. Thus, although *Cissus quadrangularis* has low reducing power, it could have high total antioxidant activity.

Based on the results obtained, it may be concluded that selected plant extract showed strong antioxidant activity, reducing power ability.



REFERENCES:

 Bandyopadhyay U, Das A and Bannerjee, RK. Reactive oxygen species: Oxygen damage and pathogenesis. Curr Sci.1999; 77(5):658-666
Halliwell B and Gutteridge JMC. Free radicals in Biology and Medicine, 3rd ed. Oxford University Press, Oxford.1999;pp. 23-27

3. Irshad M and Chaudhuri PS. Oxidant-antioxidant system: Role and significance in human body. Indian J. Exp. Biol.;2002; 40:1233-1239

4. Fang Y, Yang S and Wu G. Free radicals, antioxidants and nutrition. Nutrition,2002; 18:872-879

5. Lee J, Koo N and Min DB. Reactive oxygen species, aging and antioxidative nutraceuticals. CRFSFS.2004;3:21-33

6. Cuzzocrea S, Riley DP, Caputi AP and Salvemini D. Antioxidant therapy: A new pharmacological approach in shock, inflammation and ischemia/reperfusion injury. Pharmacol. Rev.,2001;53:135-159

7. Grice HC. Safety evaluation of butylated hydroxytoluene (BHT. in the liver, lung and gastrointestinal tract. Food Chem. Toxicol.,

1986;24:1127-1130

8. Wichi HP. Enhanced tumour development by butylated hydroxyanisole (BHA) from the prospective of effect on forestomach and oesophageal squamous epithelium. Food Chem. Toxicol., 1988;26:717-723

9. Formica JV and Regelson W. Review of the biology of quercetin and related bioflavonoids. Food Chem. Toxicol.,1995; 33: 1061-1080

10. Rice-Evans CA, Miller NJ and Paganga G. Antioxidant properties of phenolic compounds. Trends Plant Sci. Rev., 1997; 2: 152-159

11. Diallo D, Hveen B, Mahmoud Ag M, et al., An Ethnobotanical survey of herbal drugs of Gourmia District, Mali Pharm. Biol. 1999;37(1),80-91

12. Chapaganin B and Zeev W. African J. Biotech. 2005; 4(11),1209-1213

13. Kritikar and Basu. Indian Medicine Plants with illustration 3 Oriental Enterprises. 2001; 710-714

14. Singh VK, Govil JN and Singh UP. Medicinal properties of plants. 2004; International

Book Distribution Co.

15. Yildrim A, Oktay M and Bilaloglu V. The antioxidant activity of the leaves of Cydonia vulgaris. Turk. J. Med. Sci., 2001;31:23-27

16. Fejes S, Blazovics A, Lugasi A, et al. In vitro antioxidant activity of Anthriscus cerefolium L. (Hoffm.) extracts. J. Ethnopharmacol., 2000;69:259-265

17. Meir S, Kanner J, Akiri B and Hadas SP. Determination and involvement of aqueous reducing compounds in oxidative defences systems of various senescing leaves. J. Agric. Food Chem 1995;43:1813-1817

18. Yen GH and Chen HY. Antioxidant activity of various tea extracts in relation to their antimutagenicity. J. Agric. Food Chem., 1995;43: 27-32

19. Diplock AT. Will the 'good fairies' please prove to us that vitamin E lessens human degenerative diseases? Free Rad. Res.1997;27:511-532.